Behavioral/Cognitive

Frontoparietal Correlation Dynamics Reveal Interplay between Integration and Segregation during Visual Working Memory

Nicholas M. Dotson, Rodrigo F. Salazar, and Charles M. Gray

Cell Biology and Neuroscience, Montana State University, Bozeman, Montana 59717

Working memory requires large-scale cooperation among widespread cortical and subcortical brain regions. Importantly, these processes must achieve an appropriate balance between functional integration and segregation, which are thought to be mediated by task-dependent spatiotemporal patterns of correlated activity. Here, we used cross-correlation analysis to estimate the incidence, magnitude, and relative phase angle of temporally correlated activity from simultaneous local field potential recordings in a network of prefrontal and posterior parietal cortical areas in monkeys performing an oculomotor, delayed match-to-sample task. We found longrange intraparietal and frontoparietal correlations that display a bimodal distribution of relative phase values, centered near 0° and 180°, suggesting a possible basis for functional segregation among distributed networks. Both short- and long-range correlations display striking task-dependent transitions in strength and relative phase, indicating that cognitive events are accompanied by robust changes in the pattern of temporal coordination across the frontoparietal network.

Key words: electrophysiology; frontoparietal; monkey; oscillations; synchrony; working memory

Introduction

The realization that the primate cerebral cortex is composed of a large number of anatomically and functionally distinct cortical areas (Brodmann, 1909; DeYoe and Van Essen, 1988; Goldman-Rakic, 1988; Paxinos et al., 1999; Saleem and Logothetis, 2007; Van Essen et al., 2012), widely linked by weak corticocortical connections (Zeki and Shipp, 1988; Felleman and Van Essen, 1991; Markov et al., 2014), poses a well-recognized challenge to theories of perception, action, and cognition. Somehow, large populations of neurons, distributed across widespread regions of the cortex, must rapidly and selectively cooperate in a flexible manner to enable a vast repertoire of possible functions (Mesulam, 1990; Bressler, 1995; Tononi et al., 1998; Varela et al., 2001; Bressler and Menon, 2010). A large body of evidence indicates that this functional integration occurs when distributed populations of neurons transiently form large-scale networks, defined by the spatiotemporal patterns of coordination among the participating neurons (Bressler et al., 1993; Gray, 1994; Bressler,

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Correspondence should be addressed to Dr. Nicholas M. Dotson, Montana State University, 1 Lewis Hall, Bozeman, MT 59717. E-mail: nicholas.dotson@live.com.

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1995; Singer and Gray, 1995; Roelfsema et al., 1997; Varela et al., 2001; Buzsáki, 2006; Buschman and Miller, 2007; Fell and Axmacher, 2011; Siegel et al., 2012; Crowe et al., 2013). However, this functional integration must be balanced by a competing demand for segregation to avoid false associations and globally cooperative states (Zeki and Shipp, 1988; Kelso, 1995; Bressler and Kelso, 2001; Tognoli and Kelso, 2014). Segregation could occur when competing networks are inhibited (Desimone, 1998), display statistical independence (Singer and Gray, 1995; Fries, 2005), or operate at different phases or frequencies (Bressler and Kelso, 2001; Popa et al., 2009; Tognoli and Kelso, 2014).

Working memory has the hallmark features of a cognitive process, requiring both functional integration and segregation. Endogenous and/or exogenous information no longer available in the environment, must be maintained, manipulated, and integrated with behavioral goals and decisions while resisting interference from competing representations (Miller, 2013). Recently, we reported a high incidence of task-dependent and contentspecific synchronization of activity between prefrontal (PFC) and posterior parietal (PPC) cortical areas in the 12-22 Hz frequency range (Salazar et al., 2012). Although these findings demonstrate that frontoparietal synchronization reflects the integration of distributed activity, the properties of the relative phase relationships and the patterns of correlation within PFC and PPC were not analyzed. Here, we use cross-correlation analysis to estimate the incidence, magnitude, and relative phase angle among pairs of local field potential (LFP) signals sampled from 63 different intra-areal and interareal combinations within and between PFC and PPC. We find widespread patterns of task-dependent correlation, largely within the previously reported frequency band, that vary in magnitude and incidence of occurrence and fall into

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two distinct phase relationships centered near 0° and 180°. The phase relationships are dynamic and can display rapid transitions by as much as 180° that are linked to task events. These findings suggest that dynamic changes in relative phase, in addition to inhibition and statistical independence, may contribute to the interplay between functional integration and segregation of distributed networks.

Materials and Methods

Subjects. Two female rhesus macaque monkeys (Monkey A and Monkey B) were each implanted with a cranial post for head restraint, scleral search coils to track eye positions (Judge et al., 1980), and two recording chambers (Gray et al., 2007) located above the prefrontal and posterior parietal cortices (see Fig. 1*a*) (for more details see Salazar et al., 2012). All procedures were performed in accordance with National Institutes of Health guidelines and the Institutional Animal Care and Use Committee of Montana State University.

Behavioral procedures. The monkeys were trained to perform an oculomotor, delayed match-to-sample task involving a noninstructed rule switch. The results from this study are only from trials during the identity rule (see Fig. 1b). In this task, each trial is initiated with the appearance of a central fixation spot presented on a black background. The monkey, seated in a primate chair in front of a 19-inch monitor (85 Hz refresh rate, 57 cm distance), is required to maintain visual fixation within a 3° window throughout the trial until an appropriate oculomotor response is made. At a latency of 500 ms, one of three possible sample objects (size, 3°; eccentricity, 5°) is presented for 500 ms at one of three possible locations lying at the vertices of an invisible triangle. This design resulted in 9 unique sample stimuli on each session. The sample stimulus is followed by a randomized delay, ranging from 800 to 1200 ms in one set of experiments and 1 to 3 s in a second set of experiments, in which no stimulus is present. At the end of the delay period, the fixation target is extinguished and a match stimulus is presented that consists of a target and a distracter object positioned at two of the three possible locations. While the match stimulus is visible, the monkey must make a saccadic eye movement to the target object and maintain gaze on the object for 300 ms. The target object is defined by the matching rule in effect, either location or identity. After the monkey had reached criterion performance (>80% correct responses for at least 300 consecutive trials using a sliding window of 100 trials), the rule was switched without cueing the animal. Several rule switches could occur each day, but in general only one or two switches were achieved. Correct trials were rewarded with a drop of juice. To reduce biases for both the location and identities of the objects, the object locations were rotated by 180° on alternate days and each day a new set of objects was selected. All object positions and identities were presented with equal probability in a pseudorandom sequence. In Monkey B, 10% of the trials did not include stimuli and the monkey simply had to maintain fixation throughout the trial to receive a reward. The eye position signals were acquired using the scleral search coil method in Monkey A and an infrared eye-tracking system in Monkey B. Both custom software and the publicly available MonkeyLogic software (http://www.monkeylogic.net/) were used to display the stimuli and provide experimental control (Asaad and Eskandar, 2008).

Recording techniques. Broadband neuronal activity was recorded simultaneously from multiple electrodes in both the prefrontal and posterior parietal cortex (see Fig. 1*a*,*c*). The signal from each electrode was amplified (5k), filtered (1 Hz-10 kHz), digitized (30 kHz), and stored for off-line analysis. In Monkey A, an acute recording method (Gray et al., 2007) was used that involved passing 4-8 independently positioned microelectrodes (tungsten-in-glass, $0.5-1.5 \text{ M}\Omega$) through the dura on a daily basis. The horizontal separation between electrodes was at least 0.86 mm (range, 0.86-6.08 mm), and the recording depths ranged from the point of initial detection of unit activity to a maximum of 5 mm. Monkey B was implanted with two miniature mechanical microdrives containing 32 independently movable electrodes with a horizontal spacing of 1.5 mm and a maximum travel of 20 mm. The maximum recording depths were ~10 mm from the surface of the cortex. There was no attempt to identify the response properties of neurons before recording. Because of

Table 1. Sampling distribution of recording sites in both monkeys^a

Area	Sites recorded	Sites with preferred spike-filed phase	Sites with preferred phase near LFP trough
Monkey A			
6DR	0	0	0
8AD	0	0	0
8B	16	13	13
dPFC	55	51	50
vPFC	2	2	2
9L	11	9	9
PEC	45	43	41
PGM	0	0	0
PE	0	0	0
PG	38	32	32
MIP	2	2	2
LIP	5	4	4
Total	174	156	153
Monkey B			
6DR	14	14	14
8AD	44	25	21
8B	98	87	86
dPFC	90	65	62
vPFC	12	12	12
9L	0	0	0
PEC	97	95	84
PGM	2	2	1
PE	91	85	84
PG	30	22	22
MIP	66	65	63
LIP	61	57	33
Total	605	529	482

⁶The first column lists the cortical area. The second column is the total number of recording sites for each area. The third column is the number of sites with nonuniform, spike-field phase distributions, indicating a preferred phase (Rayleigh test, p < 0.05). The last column is the number of sites with nonuniform phase distributions and a preferred phase that is within $\pm 90^\circ$ (circular mean) of the LFP trough (180°). Areal abbreviations follow the nomenclature of Paxinos et al. (1999) with the exception of dPFC and vPFC: 6DR, Dorsorostral area 6; 8AD, anterodorsal area 8; 8B, area 8B; dPFC, dorsal prefrontal cortex; vPFC, ventral prefrontal cortex; 9L, lateral area 9; PEC, caudal area PE; PGM, medial area PG; PE, area PE; PG, area PE; PG, medial intraparietal area; LIP, lateral intraparietal area.

the recording technique in Monkey B, \sim 23% of signal pairs selected for analysis were recorded with electrodes at the same position during recording sessions occurring at least 24 h apart. In both sets of recordings, the reference and ground connections were tied together and connected to both of the recording chambers.

Signals were acquired in a trial-based format; the data acquisition hardware started and stopped recording at the beginning and end of each trial, respectively. In Monkey A, stop signals were received by the data acquisition hardware ~400 ms after the animal made a response. In Monkey B, stop signals were sent ~400 ms after correct responses and ~1.5 s after incorrect responses. This additional delay served to penalize the monkey after incorrect responses. The penalty delay also enabled us to analyze the intertrial interval (ITI) following error trials for Monkey B only.

The broadband signal from each electrode was bandpass filtered (1– 100 Hz) and down-sampled to 1 kHz to obtain the LFP. To extract the unit activity, the broadband signal was highpass filtered (500 Hz to 7 kHz). Spike waveforms were extracted by detecting local minima in the highpass signal that exceeded 4 and 5 SDs of the noise level in Monkey A and Monkey B, respectively. These waveforms were clustered using the KlustaKwik software package and further sorted using MClust (http://redishlab.neuroscience.umn.edu/MClust/MClust.html). Only channels with detectable unit activity were considered for further analysis to ensure that our signals were not in white matter.

Histology. When recording was completed, each monkey was euthanized (pentobarbital, 100 mg/kg i.v.) and perfused through the heart with PBS followed by a solution of 5% PFA in PBS. The brain was removed and relevant portions blocked and sunk in a solution of fixative with 30% sucrose several days before being sectioned (60 μ m) and stained for Nissl substance to identify electrode tracks (FD Neurotech-

Data selection, filtering, and spike-field analvsis. All data analysis was performed using MATLAB (MathWorks). Based on our previous analysis, we chose a frequency range for the cross-correlation analysis that closely overlapped with the content-specific patterns of frontoparietal coherence (Salazar et al., 2012). To determine the relevant upper and lower frequency limits for this analysis, we calculated the prevalence of peaks in the power spectrum of each LFP signal. The spectral power was estimated using multitaper spectral analysis (time-bandwidth product of 2, with 3 Slepian tapers, MATLAB Chronux Toolbox). The mean power during the delay period (1.2-1.8 s) was calculated for each channel using all correct trials during stable performance (80% correct). The spectra were first interpolated to a resolution of 1 Hz, smoothed (3 point moving average, 1 point step), and then investigated for peaks by finding local maxima. At each peak, the corresponding frequency was recorded. Figure 2a shows five representative mean spectra with the peaks marked by red asterisks. This approach provided us with a histogram of peak frequencies across the dataset (see Fig. 2b) that revealed the incidence of spectral power that deviates from the 1/f distribution that is commonly observed in long duration recordings of the LFP or EEG (Buzsáki and Draguhn, 2004). The presence of a peak characterizes the dominant frequency of the oscillations; however, its absence does not exclude the presence of oscillations if the underlying magnitude of the power is high. Based on the results of this analysis, we bandpass filtered the LFP signal from 8 to 25 Hz (zero phase forward and reverse digital IIR fourth-order Butterworth filter) before calculating the cross-correlograms. Figure 2c provides examples of two signals before and after filtering, illustrating that the dominant frequency band is not altered by the filtering.

The LFP signals were chosen for further analysis if the multiunit activity (MUA) recorded on the same channel had a preferred phase (Rayleigh test, p < 0.05) within $\pm 90^{\circ}$ of the LFP trough (see Fig. 3a). This selection insured that a consistent relationship was present between the unit activity and the LFP for all signals at all recording depths. To calculate the spike-field relationships, the Hilbert transform was used to estimate the instantaneous phase of the LFP at the time of each spike (bandpass filtered at 8-25 Hz). Then, a distribution of instantaneous phase values was made with these values. The circular mean of this distribution is what we refer to as the preferred phase. The Rayleigh test, which is a test for uniform circular distributions, was used to evaluate significance (p < 0.05). With these criteria, ~79% of the PFC and 75% of the PPC recordings were selected (for the cumulative phase



Figure 1. Illustration of recording locations, behavioral task, and an example dataset. *a*, Recording chamber locations in Monkey B superimposed on a photograph of the left hemisphere of a separate macaque brain. The chamber locations in Monkey A were similar. The prefrontal chambers were positioned over the dorsolateral PFC, and the parietal chambers were positioned over the posterior intraparietal sulcus. *b*, Illustration of the delayed match-to-sample task. The task involved an initial fixation period (500 ms), followed by the presentation of a sample stimulus (500 ms), consisting of one of three possible locations, a variable delay period (800 – 1200 ms or 1000 – 3000 ms) and a match stimulus, consisting of the target object and a distractor object presented at two of three possible locations. The animals successfully completed each trial by making a saccade to the target object and maintaining gaze for a minimum of 300 ms. If the distractor was chosen, the trial immediately ended. The dashed box during the match epoch is not visible to the animal and is only included to indicate the correct choice. *c*, Example ofbroadband signals recorded on a single trial from Monkey A. The black vertical lines indicate the sample onset, sample offset, and the match onset, respectively. Top and bottom voltage scales correspond to channels 1– 4 in PPC and channel 5 in PFC, respectively.



Figure 2. Summary of the spectral peak analysis. *a*, Example of the average spectra (normalized by dividing by the integral) computed for the delay period (1.2–1.8 s) for 5 channels (same channels used for the raw data examples in Fig. 1). Red asterisks indicate where peaks were detected. *b*, Distributions of peaks in the spectral power for Monkey A (left) and Monkey B (right) spanning the range of 5–45 Hz in 5 Hz bins. A total of 198 of 342 PFC signals (Monkey A = 48 of 84, Monkey B = 150 of 258) and 428 of 437 PPC signals (Monkey A = 83 of 90, Monkey B = 340 of 347) had peaks in their power spectrum. *c*, Example of broadband signals (black) recorded on a single trial from Monkey A with the bandpassed (8–25 Hz) signals overlaid (green). These are the same broadband signals used in Figure 1, and their corresponding average spectra are shown above. Gray dashed lines indicate the sample onset, sample offset, and the match onset, respectively.



Figure 3. Summary of the spike-field phase analysis. *a*, Distributions of the preferred phase (circular mean) of the MUA with respect to the LFP, for those sites passing the Rayleigh test (p < 0.05), as a function of electrode depth. In Monkey A (left), the recording depth is relative to the first detection of unit activity near the cortical surface. In Monkey B (right), the recording depth is relative to the starting position of the electrodes in the microdrive. A circular mean near 180° indicates that the MUA is concentrated near the trough, or negative phase, of the LFP (shaded region). *b*, Circular histograms of the preferred phase (circular mean) for all signals that passed the Rayleigh test with a circular mean >90° in absolute value. Inset (middle), The timing of spikes relative to the LFP oscillation, with the negative trough at $\pm 180°$. The grand circular mean is given in the bottom left of each histogram.

distributions, see Fig. 3*b*; for areal information, see Table 1).

Cross-correlation analyses. All cross-correlations were performed using the MATLAB built-in function "xcorr.m" (correlation coefficients range from -1 to 1). The average cross-correlogram (± 50 ms time lag, data down-sampled to 1 kHz) of the LFP (8-25 Hz) for each pair of signals was calculated across all correct identity trials (stable performance, >80% correct) (Salazar et al. (2012)) and all stimuli during the presample (0.1–0.5 s) and the delay period (1.2–1.8 s and 1.4–1.8 s). Because of our data acquisition technique at the time of recording, the average cross-correlograms during the ITI (0.5-0.9 s after match onset) were calculated from the incorrect trials for Monkey B only (see above for details). In order to compare the delay and ITI periods for Monkey B, the average cross-correlograms were also calculated from the incorrect trials in Monkey B during the delay period (1.4–1.8 s). The frequency of each correlogram was determined by fitting an 8 parameter generalized Gabor function (König, 1994) with the minimum squared error used as the criterion for the best fit. This approach has been widely used to quantitatively characterize the properties of auto- and cross-correlograms (e.g., Freeman, 1975; Gray and Skinner, 1988). The central peak (positive or negative) obtained from the fitted Gabor function was determined by finding the local maximum or minimum that was closest to zero time lag. The relative phase angle between the two LFPs was calculated using the time lag, polarity of the central peak, and the frequency estimated from the Gabor function. Correlograms that were near 180° out of phase had a distinct trough near zero time lag (for examples, see Fig. 4f,h). We used two criteria to identify signal pairs that were significantly correlated: (1) the absolute value of the correlation coefficient at the central peak (positive or negative) was > 0.1 ($\sim p < 10^{-20}$, randomization test), and (2) the correlation coefficient between the original correlogram and the Gabor function was \geq 0.99. To visualize the temporal aspects of these cross-correlations, time-dependent average cross-correlograms were calculated using a 200 ms window, a 50 ms time step, and a \pm 50 ms time lag (for examples, see Figs. 4a-d and 11b-d). To compare the results between task periods, we used a sign test (paired, two-sided, p < 0.05 using the Benjamini-Hochberg procedure to control the false discovery rate), which tests the hypothesis that the difference median is zero. This was used to compare the magnitude of correlation between the presample and delay periods, and the delay and ITI periods.

It could be argued that the use of a timefrequency domain analysis, such as coherence analysis, would be more useful because it avoids bandpass filtering the data. We chose to use cross-correlation analysis for several reasons. First, using the band-passed signal, we were able to directly assess the relative phase between the dominant components in the signals, rather than a combination of phase and amplitude. Second, it provided a single value for the magnitude and phase for the frequency at which the signals were maximally correlated, greatly simplifying the analysis.

ITI spectral power analysis. To determine the time course of the average power during the ITI, we first calculated the timefrequency power spectra (200 ms sliding window stepped at 50 ms) using multitaper spectral analysis (time-bandwidth product of 2, and 3 Slepian tapers, MATLAB Chronux Toolbox). Second, the maximum power at ~19 Hz was estimated relative to the match onset for each signal. The search window was delayed 200 ms after the behavioral choice to avoid activity evoked by the match stimulus. This provided us with a single value for each signal at the time after match onset where the power at ~19 Hz was highest.

Results

We recorded broadband neuronal activity from multiple electrodes in multiple areas of the PFC and the PPC in two monkeys (Monkey A and Monkey B) while the animals performed an oculomotor, delayed match-to-sample task (Fig. 1). The animals performed either an identity-matching rule or a locationmatching rule in blocks, with a noncued rule switch (see Materials and Methods). Trials were pooled across stimuli, and only trials recorded during stable performance (>80% correct) on the identity-matching rule were used for this analysis.

A total of 342 PFC and 437 PPC recordings were acquired (Table 1). Before performing any analyses, we bandpass filtered the LFP signals between 8 and 25 Hz and chose signals for further analysis when the MUA recorded on the same electrode had a preferred phase within $\pm 90^{\circ}$ of the LFP trough (Rayleigh test, p < 0.05). This ensured that the LFP signals in this frequency range had a consistent relationship with the unit activity on the same electrode. With this criterion, 79% of the PFC and 75% of the PPC recordings were selected for correlation analysis (see Materials and Methods) (Figs. 2 and 3; Table 1).

Frontoparietal correlations during working memory

To determine the patterns of functional connectivity across the frontoparietal network, we used cross-correlation analysis to estimate the incidence, magnitude, and relative phase angle between all combinations of simultaneously recorded LFPs during the delay period of the task. We found significant correlations in 60% and 48% of the signal pairs in Monkey A and Monkey B, respectively, resulting in 1644 significantly correlated signal pairs from 63 different intra-areal and interareal combinations (Table 2; for selection criteria, see Materials and Methods). Figure 4 shows four examples of the signal correlations we observed in the data from Figure 1*c*. During the task, the correlation magnitude

Table 2. Counts for all areal combinations before any selection criteria, and for four different subsets with selection criteria^a

Table 2	Continued
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Area 1	Area 2	1: Signal pairs	2: Significant	3: Significant signal pairs near 180°	4: Significan signal pairs (presample vs delav)	t 5: Significant signal pairs (ITI vs delav)
	711002	puilo	signal parts		is aciay,	(1115 deld))
Monkey A	01	0	7	٥	7	
4DEC	9L	0	/	0	/	
	9L) 15	2	0	2	
	9L 01	15	0 1	1	/	
	9L 0I	2	1	1	0	
9L 0D	9L 0D	4	Z	0	2 F	
٥D مD) 12) 10	0) 11	_
ÖD OD		13	12	0	14	
٥D	PEC	25	20	2	14	_
٥D مD		21	/	/	0	_
ÖD OD		1	1	0	1	
4DEC		1	0	0	22	
		45	22	0	52	
		74	0	0) 10	
	PEC	/4	26	2 15	18	_
	PG	53	16	15	6	_
	MIP	0	5	0	3	_
		6	5	5	3	_
VPFC	PEC	5	2	0	2	_
VPFC	PG	2	2	1	1	_
PEC	PEC	29	28	1	27	_
PEC	PG	34	23	17	22	_
PEC	MIP	4	4	0	4	_
PEC	LIP	1	0	0	0	
PG	PG	21	1/	0	18	_
PG	MIP	2	1	1	2	_
PG	LIP	2	2	0	2	_
	lotal	390	235	53	200	_
Monkey B	(00	2				
6DK	6DK	2	0	0	0	0
6DR	8AD	12	6	1	2	6
6DK	8B	31	12	3	6	8
6DK	dPFC	1/	8	2	/	6
6DK	PEC	50	2	1	1	1
6DK	PE	49	2	1	0	3
6DK	PG	10	0	0	0	0
6DK	LIP	1/	3	3	3	0
8AD	8AD	6	6	0	6	6
8AD	88	58	38	4	32	34
8AD	dPFC	36	31	0	31	26
8AD	VPFC	6	4	0	5	4
8AD		61	1/	1	13	5
SAD	PE DC	69	40	0	32	20
8AD		1/	9	U	<u>ځ</u>	/
8AD	rgw	I	1	U	1	U
8AD	MIP	9		U	0	U
0AU		10	U 10	U	0	5
δĽ	ADEC.	/5	19	5	16	18
8B 8B	dPFC	135	62	9	51	41
δĽ	VYFC	۲) ۱۵۲	0	0	/	0
δĽ		196	66	1	54	43
δĽ	PE DC	191	89	0	84	/0
δR	PG	51	30	16	1/	1/
88	PGM	2	0	0	0	0
8B	MIP	118	12	10	1	3
88	LIP	58	34	34	24	28
dPFC	dPFC	35	26	0	26	25
dPFC	vPFC	19	12	0	8	8
dPFC	PEC	128	35	0	19	13
dPFC	PE	143	74	0	62	44
dPFC	PG	42	25	7	13	13
					(Table continues.)

Area 1	Area 2	1: Signal pairs	2: Significant signal pairs	3: Significant signal pairs near 180°	4: Significant signal pairs (presample vs delay)	5: Significant signal pairs (ITI vs delay)
dPFC	PGM	2	1	0	2	2
dPFC	MIP	62	16	0	10	10
Monkey B						
dPFC	LIP	47	13	2	8	7
vPFC	vPFC	7	7	0	7	2
vPFC	PEC	5	0	0	0	0
vPFC	PE	16	12	0	9	0
vPFC	MIP	52	4	0	2	3
vPFC	LIP	16	0	0	0	2
vPFC	PGM	3	0	0	0	0
PEC	PEC	92	78	0	63	61
PEC	PE	260	188	0	158	153
PEC	PG	58	23	21	16	9
PEC	MIP	52	47	0	46	44
PEC	LIP	77	31	22	21	7
PE	PE	120	114	0	100	106
PE	PG	56	23	22	20	14
PE	MIP	51	32	0	28	32
PE	LIP	91	37	37	38	17
PG	PG	4	4	0	4	3
PG	LIP	10	7	0	7	9
MIP	MIP	96	80	0	69	75
MIP	LIP	60	0	0	2	0
LIP	LIP	15	14	0	13	13
PGM	PE	2	2	0	2	2
PGM	MIP	6	4	0	3	3
PGM	LIP	2	2	2	2	2
	Total	2953	1409	204	1154	1032

^aColumn 1, The total number of signal pairs. Column 2, The total number of signal pairs during the delay (1.2–1.8 s) that pass the cross-correlogram selection criteria. Column 3, The total number of signal pairs during the delay (1.2–1.8 s) that pass the cross-correlogram selection criteria and have a relative phase angle near 180° ($(\theta) > 90^\circ$). Column 4, The total number of signal pairs that pass the cross-correlogram selection criteria during the delay (1.4–1.8 s) and the presample (0.1–0.5 s). Column 5, The total number of signal pairs that pass the cross-correlogram selection criteria during the delay (1.4–1.8 s) and the ITI (0.5–0.9 safter match onset), during incorrect trials (Monkey B only). Areal abbreviations are the same as in Table 1.

peaks before the sample, transiently declines, and then increases again during the delay (Fig. 4a-d). These magnitude changes are accompanied by subtle shifts in the relative phase angle, especially during the response to the sample when the correlation magnitude is at its minimum. Notably, the correlation magnitudes and phase relationships are related to the cortical locations of the recording sites. In these examples, the PEC-PEC signal pair, recorded medial to the intraparietal sulcus (IPS), exhibits a strong, in-phase correlation (Fig. 4a,e), whereas the PG-PEC signal pair, recorded from opposite sides of the IPS, displays a weaker, but nearly anti-phase, correlation (Fig. 4b,f). Similar phase relationships are present in the long-distance frontoparietal signal pairs. The dPFC-PEC (Fig. 4c,g) and dPFC-PG (Fig. 4d,h) signal pairs display weaker correlations while maintaining a striking difference in relative phase.

Analysis of the incidence and magnitude of correlation with respect to spatial separation revealed distinct patterns of functional connectivity. We evaluated these effects by grouping signal pairs into six categories: (1) within PFC, (2) PPC signals medial to the IPS, (3) PPC signals lateral to the IPS, (4) PPC signals on opposite sides of the IPS, (5) PFC and medial PPC pairs, and (6) PFC and lateral PPC pairs. As shown in Table 3, the incidence and magnitude of correlation were highest among signal pairs recorded on the same side of the IPS, somewhat lower within PFC, and markedly lower for signal pairs separated by larger distances



Figure 4. Example of cross-correlation results. *a*–*d*, Average cross-correlograms as a function of time (200 ms window, 50 ms step) for four of the signal pairs shown in Figure 1*c*. Vertical black lines indicate the sample onset, sample offset, and earliest match onset, respectively. *e*–*h*, Average cross-correlograms (blue curves) calculated from correct trials during the delay period (1.2–1.8 s) and generalized Gabor functions that were fit to the data (dashed red curves). The frequency and phase angle obtained from the Gabor function are indicated in the upper right of each plot.

Table 3. The incidence and magnitude of significant correlation with respect to
cortical regions ^a

Combination	Incidence (%)	Magnitude median (25th/75th)
PFC-PFC	55	0.39 (0.27/0.51)
mPPC-mPPC	81	0.50 (0.38/0.61)
IPPC-IPPC	85	0.71 (0.54/0.79)
mPPC-IPPC	37	0.17 (0.13/0.22)
PFC-mPPC	34	0.15 (0.12/0.20)
PFC-IPPC	40	0.18 (0.14/0.22)

^oIncidence is reported as a single percentage across all measurements in each category. Magnitude is reported as the median (25th/75th percentiles). The results are grouped into six categories: PFC, mPPC-mPPC, IPPC-IPPC, mPPC-IPPC, PFC-mPPC, and PFC-IPPC. PFC, Prefrontal; mPPC, medial posterior parietal; IPPC, lateral posterior parietal.

(i.e., mediolateral parietal, frontoparietal). To assess the effect of interelectrode separation on correlation magnitude within PFC and PPC, we plotted the correlation magnitude versus distance for all pairs within PFC, and together for all pairs on the same side of the IPS. This revealed a fall off of correlation magnitude as a function of distance for electrode separations <6 mm (Fig. 5). Thus, the magnitude of correlation decreases with distance within and between adjacent areas of the PFC and PPC (Murthy and Fetz, 1996; Destexhe et al., 1999), whereas both the incidence and magnitude of correlation plateau to lower levels over distances that span widely separated areas of cortex (Roelfsema et al., 1997).

Consistent with the results in Figure 4, we found a clear bimodal distribution of relative phase values among significantly correlated pairs in both monkeys (Fig. 6). With all data combined, the distributions of relative phases were split into approximately in-phase (near 0°) and anti-phase (near 180°) modes (Fig. 6a,e). To identify the source of these distinct modes, we separated the data into PFC, PPC, and frontoparietal categories. Within PFC, the vast majority of phase values are centered near 0° in both monkeys (Fig. 6b,f). Within PPC, the relative phase distributions are distinctly bimodal in both monkeys, with the dominant mode centered near 0° and the secondary mode centered near 180° (Fig. 6c,g). These bimodal phase distributions also exist in the frontoparietal correlations in both animals (Fig. 6d,h): The dominant mode, centered near -16° in both monkeys, displays a prefrontal phase lead, as previously reported (Salazar et al., 2012), whereas the secondary mode is more broadly distributed and centered at -180° and -148° in Monkey A and Monkey B, respectively (Table 4).

To gain further insight into these processes, we subdivided the correlation analysis with respect to each intra-areal and interareal combination of signals. The magnitude, relative phase, and frequency values of the correlations are shown in Figure 7. Because the sign of the relative phase angle can be ambiguous, we report the absolute value of the relative phase. The results reveal that the patterns of correlation are anatomically specific. Within PFC, the correlations vary widely in magnitude with respect to areal combination (and thus distance) and are dominated by

in-phase relationships, with a small percentage of values scattered ~100° in Monkey B (Fig. 7a). Within PPC, signal pairs recorded on either the medial (Fig. 7b) or the lateral (Fig. 7c) side of the IPS have a broad distribution of correlation magnitudes and are highly clustered in-phase, with only one value exceeding 90°. In contrast, the long-distance correlations between signals recorded on the medial and lateral sides of the IPS, and between these areas and the PFC, are smaller in magnitude and exhibit a clear bimodal distribution of phase values (Fig. 7d-f). Signal pairs recorded on opposite sides of the IPS are dominated, with some exceptions, by near anti-phase relationships (Fig. 7d). Signal pairs recorded in PFC and the medial side of the IPS (Fig. 7e) are dominated by in-phase relationships, with the exception of signals sampled from areas 8B and MIP in Monkey B that are almost entirely out-of-phase. In contrast, signal pairs recorded in PFC and the lateral side of the IPS (Fig. 7f) exhibit both unimodal and bimodal phase distributions that are anatomically specific. In Monkey A, all signal pairs, except one, have phase values >90°. In Monkey B, correlations between prefrontal and lateral parietal signals are either exclusively out-of-phase (8B-LIP), exclusively in-phase (8AD-PG), or bimodally distributed (8B-PG, dPFC-PG, dPFC-LIP). Figure 7 (bottom row of plots) shows the corresponding frequencies of correlation derived from the fitted functions. The frequencies vary within the band of interest, display some subtle difference with respect to areal combination, and differ between the two animals. Moreover, there is no apparent difference in correlation magnitudes between the two animals (also see Fig. 5), even though the dominant frequencies are different. Clear examples of this can be seen by examining the distributions for the dPFC-PEC and PEC-PEC pairs. These subtle differences in frequency may provide another means for balancing integration and segregation.

In addition to the abundance of correlated interactions, we also found a number of interareal pairings with significant correlation occurring $\leq 10\%$ of the time (Fig. 8; Table 2). Although this effect might be related to small sample sizes (e.g., 6DR-PG) and thus reflect an experimental bias, other interareal combinations have ≥ 50 signal pairs and show a very low probability of correlation. For example, none of the cross-correlograms computed from the 60 MIP-LIP signal pairs in Monkey B reached significance. This indicates that these and other areas are functioning largely independently under the conditions of this experiment.

To further illustrate the specificity of these interactions, Figure 9 shows the correlation patterns obtained from a single

recording session in Monkey B. Within PFC, the signal correlations have values <0.5 and, with one exception, display relative phase values $<45^{\circ}$ (Fig. 9c). The intraparietal signals exhibit stronger correlations having relative phase values <45° when the signals are recorded on the same sides of the IPS, and weaker correlations with relative phases >135° for recordings on opposite sides of the IPS (Fig. 9d). The frontoparietal correlations are weaker and display relative phase values that are clustered into two groups (Fig. 9*e*): those $<45^{\circ}$ and those $>135^{\circ}$. This segregation of phases, as in Figure 4, displays some anatomical boundaries. Signals in area 8B exhibit near anti-phase relationships with signals recorded in areas LIP and MIP, even though the signals in LIP and MIP are statistically independent (Fig. 8). The remaining prefrontal signals in this example in areas 8AD, dPFC, and vPFC have in-phase relationships with parietal signals recorded in areas that are medial to the IPS but are statistically independent from signals recorded lateral to the IPS, even though significant antiphase correlations occur between signals that span the IPS. These patterns of correlation suggest that in-phase and anti-phase relationships are established at different times among different sites in the network.

Task-dependent correlation patterns

Next, we examined the task dependence of frontoparietal correlations by comparing the presample period (0.1-0.5 s) with a window of the same size in the delay period that is locked to the first possible match (1.4-1.8 s). Signal pairs were selected using the same criteria, but for both task periods, resulting in a new set of signal pairs (Table 2). We tested whether the correlation increased or decreased between the presample and delay periods for each areal combination (sign test, paired, two-sided) and plotted the difference distributions (minimum of 5 signal pairs) pooled across both animals (Fig. 10). Several trends emerged from this analysis. Within PFC, a small, but significant, increase in correlation occurred during the delay for areal combinations 8B-8B, 8B-8AD, and 8B-dPFC. Within PPC, correlations displayed a distinct pattern of changes: areas medial to the IPS displayed a robust increase in correlation during the delay (Fig. 10b), those lateral to the IPS showed no change (Fig. 10c), and areas separated by the IPS tended to decrease during the delay, but only

Figure 5. Summary of the relationship between distance and correlation magnitude during the delay period (1.2–1.8 s). *a*, All signal pairs recorded in PFC (black squares) and from the same side of the IPS in PPC (red squares) for both animals. *b*, *c*, Signal pairs recorded in PFC and PPC from Monkey A. *d*, *e*, Signal pairs recorded in PFC and PPC from Monkey B. Green lines indicate the least-squares fit: correlation values (*r*) are provided in the upper right of each plot, along with *p* values for testing the hypothesis of no correlation, which is computed by transforming the correlation to create a test statistic having *n*-2 degrees of freedom.





Figure 6. Distributions of the relative phase angles during the delay period (1.2–1.8 s) for different combinations of signal pairs recorded from Monkey A (left column) and Monkey B (right). *a*–*d*, All, PFC, PPC, and PFC-PPC signal pairs for Monkey A. *e*–*h*, All, PFC, PPC, and PFC-PPC signal pairs for Monkey B. The yellow bars represent the circular mean \pm 1 circular SD for each identifiable mode.

Table 4. Data accompanying Figure 6^a

Combination	Monkey A	Monkey A				Monkey B			
	Count (in-phase)	In-phase (mean \pm SD)	Count (anti-phase)	Anti-phase (mean \pm SD)	Count (in-phase)	In-phase (mean \pm SD)	Count (anti-phase)	Anti-phase (mean \pm SD)	
All	182	-7.3 ± 20.7	53	181.7 ± 30.0	1205	-5 ± 19.1	204	199.5 ± 32.1	
PFC	67	-0.2 ± 12.2	0	NA	213	-5.5 ± 18.9	24	NA	
PPC	56	-8.0 ± 22.5	19	185.2 ± 24.4	582	6.3 ± 13.5	104	185.3 ± 18.1	
PFC-PPC	59	-15.2 ± 23.6	34	-179.5 ± 32.6	410	-15.9 ± 18.7	76	-148.0 ± 30.0	

^aWith all data combined, and with the data subdivided into PFC, PPC, and PFC-PPC categories, the distributions of relative phase angles were split into approximately in-phase and anti-phase modes, with a dividing line centered at ±90°. Here we report the counts and the circular mean ± SD for the in-phase and out-of-phase signal pairs. No out-of-phase data are reported for the PFC combination because of the lack of any distinct mode. NA, Not applicable.



Figure 7. Distributions of the peak correlation coefficients (top row), absolute values of the relative phase angles (middle row), and the frequency at which the signals are correlated (bottom row) during the delay period (1.2–1.8 s) for signal pairs recorded in PFC (*a*), mPPC (*b*), IPPC (*c*), mPPC-IPPC (*d*), PFC-mPPC (*e*), and PFC-IPPC (*f*) for Monkey A (red) and Monkey B (blue). Only those areal combinations with five or more observations for at least one of the animals are included in the table. *d*–*f*, Light gray shading represents areal combinations with a high percentage of phase values between 90° and 180°. Data points were jittered horizontally for better visualization.

significantly for PE-LIP (Fig. 10*d*). A related pattern of changes was apparent in the frontoparietal correlations. Significant increases occurred between dPFC and medial parietal areas PEC, PE, and MIP (Fig. 10*e*) and between prefrontal areas dPFC and 8B and lateral parietal areas PG and LIP (Fig. 10*f*). In addition, we found substantial task-dependent changes in relative phase (Fig. 10, bottom plots). Shifts in relative phase between the two task periods ranged from 0° to 45°, with the largest shifts

occurring most often between frontoparietal areal combinations (Fig. 10e, f).

To further explore the potential for these activity patterns to reorganize, we compared the delay period of the task (1.4-1.8 s)to the ITI shortly after the match period on error trials in Monkey B (For technical reasons, the ITI period was only recorded after incorrect trials in Monkey B; for further explanation, see Materials and Methods.) During the ITI of error trials, we observed a



Figure 8. Summary of the counts and incidence of significance for all areal combinations with ≥10 signal pairs. *a*, Counts for the number of signal pairs (blue bars) and the number of significant signal pairs (green bars) during the delay period (1.2–1.8 s). *b*, Percentage of significant signal pairs.



Figure 9. Example of a single recording session from Monkey B. *a*, Illustration of the recording chamber locations. The names of the sulci are abbreviated as follows: PS, principle sulcus; Arc., arcuate sulcus; and IPS, intraparietal sulcus. *b*, Color scale for the relative phase angles (absolute value). The magnitude of correlation is indicated by the line thickness. *c*-*e*, Intraareal and interareal phase angles and magnitudes illustrated on schematic drawings of the PFC (left) and PPC (right). Because of the chamber orientation and the curvature of the sulci, the recording locations do not always appear on the appropriate side of the sulcus.

strong oscillatory response, visible in the LFP, which showed a marked change in relative phase among the signals (Fig. 11a). Analysis of the peak amplitude of these oscillatory bursts revealed that they occurred earlier in PPC (671 \pm 96 ms, after match onset) than PFC (769 \pm 137 ms, after match onset) (Wilcoxon rank sum test, $p \ll 0.001$). Using this timing information, we centered the ITI analysis epoch over the peak amplitudes of the bursts (0.5–0.9 s after match onset; see Materials and Methods). Signal pairs were once again subjected to the fitting criteria for both task periods, resulting in a slightly different set of pairs (Table 2). Figure 11b, c shows the time course of the trialaveraged cross-correlation for two of the signal pairs from Figure 11a. In these examples, the relative phase is near 180° for the duration of the task and then abruptly flips to an in-phase relationship during the ITI. This effect, accompanied by an increase in correlation strength, occurs between PFC and PPC signals (Fig. 11b) and between the parietal signals on opposite sides of the IPS (Fig. 11c), demonstrating its widespread nature. Figure 11d shows the third frontoparietal pair, which shows a weak in-phase correlation before the sample, is largely uncorrelated during the delay, and then is highly correlated during the ITI, with no change in relative phase.

We quantified these task-dependent changes in the full dataset using the same method shown in Figure 10. Figure 12a-f (top plots) shows the difference distributions in correlation magnitude for each areal combination (outliers not shown). In general, correlation strength increased during the ITI, but the range of change was much larger than that seen during the task, indicating widespread activation and, with few exceptions, increased synchronization across the frontoparietal network. Correlation strength increased within and between several combinations of prefrontal areas 8AD, 8B, dPFC, and vPFC (Fig. 12*a*), among parietal areas medial to the IPS (Fig. 12b), and somewhat heterogeneously among the other prefrontal and parietal areas (Fig. 12c-f). We also found a striking change in relative phase relation-



Figure 10. Distributions of the difference in correlation coefficients (top row) and relative phase angles (bottom row) between the delay period (1.4-1.8 s) and the presample (0.1-0.5 s) for signal pairs recorded in PFC (*a*), mPPC (*b*), IPPC (*c*), mPPC-IPPC (*d*), PFC-mPPC (*e*), and PFC-IPPC (*f*). Data are combined across animals, and only those areal combinations with five or more observations are included in the table. Top row, Positive values indicate that the correlation is higher during the delay period. Red asterisks indicate that the hypothesis that the difference median is zero has been rejected (sign test, paired, two-sided, *p < 0.05 using the Benjamini–Hochberg procedure to control the false discovery rate). Bottom row, Real values of the relative phase angles were used to calculate the difference between task periods, but for visualization purposes, the relative phase angle difference is presented as an absolute value. Also for visualization purposes, data points were jittered horizontally.

ships that occurred predominantly among signals separated by large distances (Fig. 12a-f, bottom plots). Whereas the changes in phase during the task were all $<50^{\circ}$ and often much closer to 0° (Fig. 10), during the ITI following error trials we found numerous changes in phase exceeding 90°. Also, changes in correlation strength and relative phase were not obligatorily coupled. We found multiple areal combinations where correlation strength changed while relative phase remained stable, and vice versa.

A potential concern is that the patterns of correlation during the delay period may differ substantially between correct and incorrect trials, leading to the effects reported in Figures 11 and 12. We examined this possibility and found no significant difference in correlation magnitude across the population of areal combinations (paired sign test, p = 0.18), and the mean difference in relative phase angle was <4.5°.

Discussion

Our analysis reveals widespread, task-dependent temporal correlations of LFP signals in the 8–25 Hz frequency range within and between multiple areas of PFC and PPC during visual working memory. Correlation magnitude decreases with distance within and between adjacent cortical areas, whereas both the incidence and magnitude of correlation plateau to lower levels over larger distances. Also, some long-distance areal combinations show little or no evidence of correlation. Analysis of the relative phase angles revealed a clear bimodal distribution among signals recorded in widely separated cortical areas in both monkeys. Although this is not the first report of anti-phase correlations (Gregoriou et al., 2009), it is, to our knowledge, the first report of simultaneous in-phase and anti-phase correlations occurring in the same frequency range. Overall, our findings agree with what might be predicted from cortical anatomy (Markov et al., 2014). However, the bimodal distribution of relative phase, and the task dependence of correlation magnitude and phase, indicate that the interactions are not hard coded, but simply constrained by anatomy. Thus, our findings demonstrate that the correlation structure across the frontoparietal network can rapidly reorganize in a manner that is linked to behavior, indicating that largescale functional integration and segregation are dynamically regulated.

Methodological considerations

Because we used monopolar recording methods, volumeconducted signals may have contributed to our estimates of correlation. However, several factors suggest that volume conduction effects are minimal. First, intracortical microelectrodes are known to record highly local LFPs (Gray and Singer, 1989; Katzner et al., 2009; Wang and Roe, 2012; Xing et al., 2012) that exhibit minimal volume conduction (Engel et al., 1990). Thus, given that most of our recordings were separated by >2 mm, we expect volume conduction effects to be small. Second, if volume conduction were contributing significantly to our findings, we would not expect to find task- and time-dependent phase differences between sites (Salazar et al., 2012). Third, the presence of anti-phase correlations and the absence of correlations among some signals spanning the IPS indicate that even closely spaced electrodes, lying in separate cortical areas, record distinct neural signals. Together, this evidence argues against a significant contribution of volume conduction at all spatial scales measured.

It could be argued that the phase differences we observe depend on the cortical depth of our recordings. Cortical LFP signals exhibit phase differences, and polarity reversals, as a function of depth (Mitzdorf and Singer, 1978; Murthy and Fetz, 1996; Bollimunta et al., 2008) that could lead to dispersion of our measured phase values. Several of our findings, however, indicate that sampling bias across a cortical dipole field cannot account for the anti-phase relationships we observe. For example, in order for laminar sampling bias to account for the antiphase correlations between areas PE and LIP (Fig. 7*d*), and the exclusively in-phase correlations within areas PE (Fig. 7b) and LIP (Fig. 7*c*), it would require that signal measurements were made on one side of the dipole (i.e., superficial layers) in one area and the opposite side of the dipole (i.e., deep layers) in the other area. Similar arguments apply to all the paired recordings made on the same side of the IPS (Fig. 7b,c). Those phase distributions are somewhat dispersed, but there is no evidence of anti-phase relationships as might be expected from a dipole field. Furthermore, any doubt that the anti-phase relationships are functional, and not due to a sampling bias across a cortical dipole, is dispelled by the rapid and pronounced changes in relative phase following error trials (Figs. 11 and 12).

Functional role of frontoparietal phase relationships

How then should we interpret the bimodal distribution of intraparietal and frontoparietal phase relationships in Figures 6 and 7? The simplest interpretation is that during the task two large-scale frontoparietal networks are established:

one involving in-phase coupling ($|\theta| < 90^{\circ}$) between medial parietal and prefrontal areas, and the other involving anti-phase coupling ($|\theta| > 90^{\circ}$) between lateral parietal and prefrontal areas. However, this interpretation may be too simplistic. For instance, signals in areas 8B and MIP display largely anti-phase correlations, whereas signals in areas PG and LIP display differing combinations of in-phase and anti-phase correlations with prefrontal areas 8AD, 8B, and dPFC. There are also a small percentage of intraprefrontal correlations with relative phase values near or >90°. These data suggest that the phase of correlated activity reflects specific patterns of coupling among many of the prefrontal and parietal areas with a strong bias for anti-phase coupling across the intraparietal sulcus.

The question also arises as to how to interpret the specific relative phase values. Presumably in-phase and anti-phase coupling have differing functional consequences. However, given the large distances separating the recorded sites, with transmission delays ranging from \sim 5 to 10 ms, there is no simple functional interpretation. One possibility is that the anti-phase correlations occurring across the IPS, and between certain frontoparietal areal



Figure 11. Example of the ITI phase flips during a single trial and averaged over all incorrect trials for a single recording session from Monkey B. *a*, One PFC (red) and two PPC channels (green and blue) recorded simultaneously. Data are bandpass filtered from 8 to 25 Hz. Gray shading represents the time window used during the delay and ITI periods. *b*–*d*, Average correlograms as a function of time using a 200 ms sliding window with a 50 ms step for the 8B-LIP, PEC-LIP, and 8B-PEC signals shown above. Vertical black lines indicate the sample onset, sample offset, and earliest match onset, respectively. The dashed vertical blue and green lines indicate the earliest start time and latest end time for the ITI analysis window, respectively.

combinations, as well as the low incidence of correlation between certain areal combinations, such as MIP and LIP, reflect functional segregation that is imposed by the oculomotor and visual mnemonic nature of the task. This seems likely, given the differing functional properties of neurons in these various prefrontal (Petrides, 2005; Lanzilotto et al., 2013) and posterior parietal (Breveglieri et al., 2008; Andersen and Cui 2009) cortical areas.

Other aspects of the frontoparietal phase relationships require explanation. The data in Figure 4 suggest that in-phase and antiphase coupling between PFC and medial and lateral parietal areas occur simultaneously, whereas the results in Figure 9 suggest some degree of independence in the frontoparietal networks. However, the interpretation of these findings must be gauged by the fact that the median value of the long-distance correlation coefficients is <0.2. This indicates that most of the time the signals in widely separated areas of the frontoparietal network are independent and that coupling between distant areas is likely to be highly transient (Gray et al., 1992). Thus, analysis of the temporal properties of the correlations on single trials will be necessary to reveal the underlying spatiotemporal organization.



Figure 12. Distributions of the difference in correlation coefficients (top row) and relative phase angles (bottom row) between the ITI period (0.5-0.9 s locked to match onset) and the delay period (1.4-1.8 s) for signal pairs recorded in PFC (a), nPPC (b), IPPC (c), mPPC (d), PFC-IPPC (d), and PFC-IPPC (f), during incorrect trials. Data are only from Monkey B, and only those areal combinations with five or more observations are included in the table. Top row, Positive values indicate that the correlation is higher during the ITI period. Red asterisks indicate that the hypothesis that the difference median is zero has been rejected (sign test, paired, two-sided, *p < 0.05 using the Benjamini–Hochberg procedure to control the false discovery rate). Bottom row, Real values of the relative phase angles were used to calculate the difference between task periods, but for visualization purposes, the relative phase angle difference is presented as an absolute value. Also for visualization purposes, data points were jittered horizontally.

Our previous findings that frontoparietal coherence in the 12–22 Hz frequency range is content specific (Salazar et al., 2012) imply that the corresponding phase relationships may also contribute to short-term representations. Determining if and how this occurs will require a more extensive analysis than we have conducted here. The relative phase and frequency of coherent activity may vary systematically with different aspects of working memory, such as content, load, and task dependence, thereby adding further dimensions to the representations. Another possibility, discussed above, may be that the bimodal distribution of relative phases provides a mechanism for task-dependent segregation of activity into distinct networks, thereby reducing potential interference among competing representations. This could be tested by switching from a saccade-based task to one in which choices are based on varying combinations of reach and saccade. In such an experiment, the phase relationships between different components of the frontoparietal network may change as the task switches from choices based on saccades to reach to a combination of the two (Dean et al., 2012).

Our finding of abrupt and marked changes in frontoparietal correlations following error trials (Figs. 11 and 12) suggests a link to the cognitive processing of errors (Larson et al., 2014). The widely distributed and highly synchronized bursts of activity may correspond to the well-documented error-related negativity (Falkenstein et al., 1991; Gehring et al., 1993) and reflect processes related to reinforcement learning (Holroyd and Coles, 2002) mediated by midbrain dopamine systems (Schultz et al., 1997). However, the present data are limited, and further experiments are required to test these hypotheses.

Coordination in large-scale networks

The phase of rhythmic correlation has been postulated to be a critical variable controlling the selective transmission, or communication, of signals between separate neuronal populations (Fries, 2005; Womelsdorf et al., 2007). Communication between neuronal groups is thought to be most effective when they oscillate at the same frequency and are synchronized at relative phases near 0°. This model is consistent with our understanding of synaptic integration (Abeles, 1982; König et al., 1995; Azouz and Gray, 2000, 2003) and is supported by experimental evidence (Engel et al., 1991a, b; Roelfsema et al., 1997; Salazar et al., 2004). Other studies have found substantial phase differences between distant neuronal populations (Gregoriou et al., 2009; Liebe et al., 2012) and suggested that these effects reflect directional influences mediated through relatively fixed conduction delays. However, neither of these perspectives is fully consistent with our findings. We find robust correlations in a common frequency range over a wide span of spatial separations that exhibit a bimodal distribution of relative phases. Moreover, the phase relations are not fixed. They vary during the task and exhibit large transitions during the intertrial interval following error trials. A potentially more favorable interpretation of these findings is that these dynamics are a product of global cooperation across the brain and that changes in correlation magnitude and phase reflect shifts in global coordination states that are not predictable on the basis of expected transmission delays (Bressler and Kelso, 2001; Tognoli and Kelso, 2014).

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